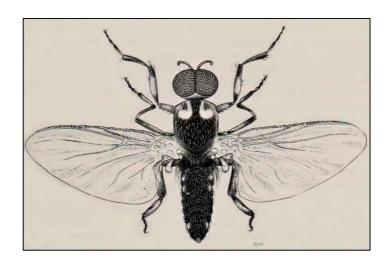
The Simuliid Bulletin

Number 57

January 2022





THE SIMULIID BULLETIN

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<u>Cover Image:</u> Male of Simulium (Odagmia) pontina depicted in Rivosecchi(1978)

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From the Editor

It is already two years and four Simuliid Bulletins ago that a virus, which has nothing in common with black flies but affects everything worldwide, started to reduce our in-person contacts. We were informing you mostly about postponed and cancelled meetings. In the beginning, many of us probably did not believe it would last long. Although the practical details on forthcoming events are still missing, there are indications that the situation is finally improving. If the lockdowns and travel restrictions are lifted, we will have the opportunity to meet at the IXth International Simuliidae Symposium in Morocco in September 2022.

Until then, we want to encourage you to share your papers, research ideas, or comments on the pages of the Simuliid Bulletin.

Tatiana Kúdelová, Editor

FORTHCOMING MEETINGS

The IX International Simuliidae Symposium, Chefchaouen, Morocco

Dear friends and colleagues,

If the pandemic situation improves, The IXth International Simuliidae Symposium is planned for

September 2022

You are cordially invited by Boutaïna Belqat and the other members of the organizing committee to **Chefchaouen, Morocco** to meet with colleagues and share your research ideas on black flies.

The exact dates, prices, and further details will be available at the Simuliid Bulletin website.

MEETING REPORTS

10th EMCA Conference: "New insights into mosquito and blackfly control"

David López-Peña

Universitat de València, Institut Cavanilles de Biodiversitat i Biologa Evolutiva, Laboratory of Entomology and Pest Control, C/ Catedrático José Beltrán, 2, 46980 Paterna (Valencia), Spain

The most recent European Mosquito Control Association (EMCA) International Conference was held in Vienna, Austria, between the 3rd and 7th of October. It was particularly relevant because it was the first time that blackfly research was included as a central topic of the conference, as indicated in the conference theme: "New insights into mosquito and blackfly control".

The session "Blackflies in Europe: where are we, where do we go?" on Tuesday 5th October consisted of several pertinent talks from blackfly researchers. The keynote was delivered by R. Bernotiene from the Nature Research Centre, Lithuania and A. Ignjatović Cupina from University of Novi Sad, Serbia, and was titled Blackfly pest species and their control: experience from Lithuania and Serbia. Following this were three additional presentations. The first one, DNA barcoding, identification, and taxonomy of blackflies in Europe, was based on work by M. Kúdela, T. Kúdelová, B. Bujačková, I. Lužáková and S. Krčmárik from Comenius University in Bratislava, Department of Zoology, Slovak Republic. The second presentation was Recovering blackfly (Diptera: Simuiidae) sample records for Tormes river basin in Spain by D. López-Peña¹, J.D. Asís-Pardo², M. Portillo-Rubio² and R. Jiménez-Peydró¹ from ¹Universitat de València (Estudi General), Institut Cavanilles de Biodiversitat i Biologa Evolutiva, Laboratory of Entomology and Pest Control, Spain, and ²Universidad de Salamanca, Facultad de Biología, Área de Zoología, Departamento de Biología Animal, Parasitología, Ecología, Edafología y Química Agrícola, Campus Miguel de Unamuno, Spain. The final talk was, Blackflies (Diptera: Simuliidae) and their parasitic watermites (Acari: Hydrachnidia), eastern Spain by D. López-Peña¹, R. Gerecke² and R. Jiménez-Peydro¹ from ¹(see above) and ²University of Tübingen,

Department of Evolution and Ecology, Tübingen, Germany.

Details about all of the mentioned presentations are available at the special Supplement 1 of the Xth International EMCA

Conference of the Journal of the European Mosquito Control Association volume 39, ISSN 2054-930X

(https://www.wageningenacademic.com/doi/epdf/10.52004/jemca2021.s1).







The attending audience was highly interested in all of the presentations and asked several questions. The attendees also took advantage of the more relaxed atmosphere during the coffee breaks to make conversation with members of the blackfly contingent and ask further questions about their research.



The conference programme also included an excursion, which consisted of two different parts. The first one was a mosquito control demonstration. Attendees witnessed the aerial application of larvicide using a helicopter and a drone. The larvicide consisted of Bacillus thuringiensis subspecies israelensis (Bti) in granulate format

to control the larval populations of mosquito pest species. All of it was performed by Heli-Austria (www.heli-austria.at). The experience was enjoyable and educational because it allowed the attendees to witness where and how the product is loaded, the suitable areas for that purpose, and the way the manned and unmanned aircraft carry out the product application (see the attached picture).

The second part involved a visit to the Palace of Schloss Hof (Schlosshof). The tour guides kindly explained the history of this amazing building. We were shown the astonishing rooms, the cosy and bright interior patios, the geometrically-designed and colourful exterior gardens, and a school farm and petting zoo in which Jezersko-Solčava sheep with four horns and a white baroque donkey stood out among other rare breeds. The guide highlighted that the 'White Baroque Donkey' (*Equus asinus f. africanus* Linnaeus, 1758) is an exceptionally rare breed of which there are only a few hundred individuals left. With a height of 105-120 cm, a weight of approximately 200 kg and a life expectancy of 40 years, the white baroque donkey was originally bred in the 16th century. During that period, the white colour was considered very special. Besides that, they are characterized by having a slight yellow tint called "cremello", as can be seen in the picture.







Eventually, the attendees were welcomed to a living room of great dimensions where the buffet gala dinner took place. This was accompanied by a live concert of Austrian traditional folk music performed by musicians in traditional attire. For those wanting extra information about the palace, visit the following link https://www.schlosshof.at/en/.

SCIENTIFIC PAPERS

Intriguing Genes: Expressed Sequences from the Simulium vittatum-tribulatum complex. IV. You are what you eat.

Charles L. Brockhouse*¹, Julia A. Brockhouse^{1,2}, and Alexie Papanicolaou³

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- 3: Hawkesbury Institute for the Environment, Western Sydney University, Sydney, Australia

Introduction

The S. vittatum-tribulatum transcriptome on which these "Intriguing Genes" reports are based consist of assemblies of approximately 28,000 S. vittatum expressed gene sequences ("contigs"), 38,000 S. tribulatum contigs, and 39,000 contigs coassembled from both species. The vittatum material was provided by the UGA Entomology colony, while the tribulatum was collected from a stream south of Omaha, Nebraska. Each species was further subdivided by sex and developmental stage to allow comparison of the expression levels during development. RNA was extracted from different stages, reversed transcribed to cDNA, and sequenced. The sequence reads were then assembled into longer sequences ("contigs") that represent expressed genes, not merely genes present in the organism. The unassembled reads from each stage can then be mapped onto each contig to give a relative measure of how much each contig is expressed in the different stages. More detailed background on the project was given in previous Bulletin issues.

We found 192 contigs expressed during larval stages, but not in adults, while 619 are expressed during all adult stages but not in the larvae. A full analysis of the differentially expressed genes is included in our upcoming transcriptome paper, but we previously presented one particular group in our NABFA and International

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Simuliidae Symposium presentations: sequences derived from the diet of the larvae. Blackfly larvae are well established as efficient filter feeders, but also grazers. Here we give genomic evidence of ingestion of rodent tissue by blackfly larvae, uncovered during manual curation of larval-specific genes.

Results:

During manual curation, three larval-specific sequences were found to have no match to Dipteran genomes in standard BLAST searches (Altschul et al., 1990), but had multiple exact matches to mouse sequences, and looser matches against human and other mammalian sequences. Two of the sequences were expressed in both sexes and one was confined to females. Both show a relatively low expression (measured by FPKM value; Fragments Per Kilobase of transcript per Million mapped reads).

1) The mouse lipocalin 2 gene:

The first two contigs matched different portions of a mouse lipocalin gene and shared the same expression profile suggesting that they are sequenced fragments of the same mRNA molecules.

>c178935_g1_i1|m.73902 matches rodent lipocalin 2gene TTCCGGGGCAGGTACGTTGTGGGCCTGGCAGGCAATGCGGTCCA GAAAAAAACAGAAGGCAGCTTTACGATGTACAG CACCATCTATGAGCTACAAGAGAACAATAGCTACAATGTCACCTCCATC CTGGTCAGGACCAGGACCAGGGCTGTCGCT ACTGGATCAGAACATTTGTTCCAAGCTCCAGGGCTGGCCAGTTCACTCT

GGGAAATATGCACAGGTATCCTCAGGTACAG
AGCTACAATGTGCAAGTGGCCACCACGGACTACAACCAGTTCGCCATG
GTATTTTTCCGAAAGACTTCTGAAAACAAGCA
ATACTTCAAAATTACCCTGTATGGAAGAACCAAGGAGCTGTCCCCTGAA
CTGAAGGAACGTTTCACCCGCTTTGCCAAGT
CTCTGGGCCTCAAGGACGACAACATCATCTTCTCTGTCCCCACCGACC
AATGCATTGACAACTGA

FPKM in female larvae: 0.79 FPKM in male larvae: 1.8 FPKM all other stages: 0



Fig. 1. Alignment of a *S. tribulatum* larval expressed sequences with a mouse lipocalin 2 gene. The two sequences are exact matches. (NCBI Megablast output)

2) The mouse aldolase gene:

This mouse sequence was confined to female larvae.

>c40815_g1_i1|m.1135 female larvae only mouse aldolase AGCATCTGCCAGCAGAATGGCATTGTACCCATTGTGGAGCCTGAAATTC TCCCTGATGGGGACCATGACTTGAAGCGCTG CCAGTATGTTACTGAGAAGGTCCTGGCGGCTGTCTACAAGGCTCTGAG CGACCACCATGTCTATCTGGAAGGCACATTGC

TGAAGCCCAACATGGTCACCCCTGGCCATGCTTGCACCCAGAAATTTTC CAATGAGGAGATTGCCATGGCAACGGTCACA GCACTTCGTCGCACAGTGCCCCCTGCTGTCACTGGGGTCACTTTCCTG TCTGGAGGGCAGAGTGAGGAAGAG

FPKM female larvae: 1.13. FPKM all other stages: 0

Homo sapiens lipocalin 2 (LCN2), mRNA

Sequence ID: NM 005564.5 Length: 820 Number of Matches: 1

Range 1	: 212	to 670 GenBank	Graphics		▼ Next Match ▲	Previous Mato
Score 243 bits(269)		Expect 3e-61	Identities 337/467(72%)	Gaps 10/467(2%)	Strand Plus/Plus	
243 DIC	(209)	36-01	337/467(72%)	10/46/(2%)	Plus/Plus	
Query	1	TTCCGGGGCAGGT	GGTACGTTGTGGGCCTGG	CAGGCAATGCGGTCCA	Gaaaaaa-aCAGA	59
bjct	212	ttccaggggaagt	ĠĠŦĀŦĠŦĠĠŦĀĠĠĊĊŦĠĠŒ	CAGGGAATGCAATTCT	cagagaagacaaa	271
uery	60	AGGCAGCTTTACG	ATGTACAGCACCATCTAT	GAGCTACAAGAGAACA	ATAGCTACAATGT	119
bjct	272	GACCCGCAAAA-G	AtGTATGCCACCATCTATO	GAGCTGAAAGAAGACA	AGAGCTACAATGT	330
uery	120	CACCTCCATCCTG	GTCAGGGACCAGGACCAG	GCTGTCGCTACTGGA	TCAGAACATTTGT	179
bjct	331	çyççtççetççtê	TTTAGGAAAAAGAAG	tgtgactactgga	tcaggacttttgt	384
uery	180	TCCAAGCTCCAGG	GCTGGCCAGTTCACTCTG	GGAAATATGCACAGGT	ATCCTCAGGTACA	239
bjct	385	tccagettgccag	cçcqqçqqqttçqçqçtqq	gcaacattaagagtt	Accctggattaac	444
uery	240	GAGCTACAATGTG	CAAGTGGCCACCACGGACT	TACAACCAGTTCGCCA	TGGTATTTTTC-C	298
bjct	445	gygttycctcgtc	CGAGTGGTGAGCACCAAC	LYCYYCCY GCALGCLY	tggtgttcttcaa	504
uery	299	GAAAGACTTCTGA	AAACAAGCAATACTTCAAA	AATTACCCTGTATGGA	AGAACCAAGGAGC	358
bjct	505	gyyge-14454cy	AAACAGGGAGTACTTCAAG	gatcaccctctacggg	AGAACCAAGGAGC	563
uery	359	TGTCCCCTGAACT	GAAGGAACGTTTCACCCGG	CTTTGCCAAGTCTCTG	GGCCTCAAGGACG	418
bjct	564	tgacttcggaact	AAAGGAGAACTTCATCCGG	cttcrccqqatctctd	GGCCTCCCTGAAA	623
uery)	419	ACAACATCATCTT	CTCTGTCCCCACCGACCA	ATGCATTGACAACTGA	465	
Sbjct	624	Yccycy4ce4c44	ccctetcccaatceacca	stgratcgacggctga	670 2	

Fig. 2. Alignment of the *S. tribulatum* putative mouse sequence with the human lipocalin 2 gene, showing multiple mismatches. The blackfly sequence does originate from human contamination. (NCBI Megablast output)

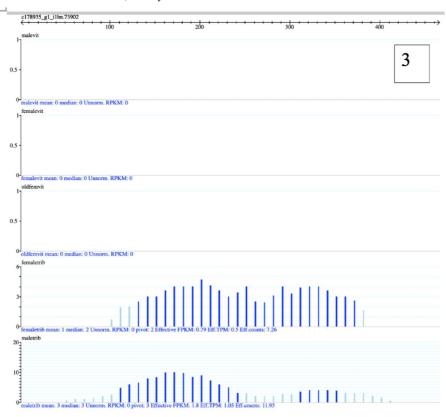


Fig. 3. Expression profiles of 2 fragments of a mouse lipocalin gene. The top 3 panels represent the adult stage; larval females and males are shown in the bottom 2 panels. The mRNA for this sequence was present only in the larval samples.

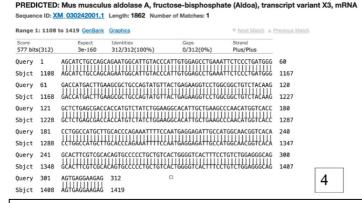


Fig. 4. Sequence alignment between blackfly contig and mouse aldolase sequence (NCBI Megablast output)

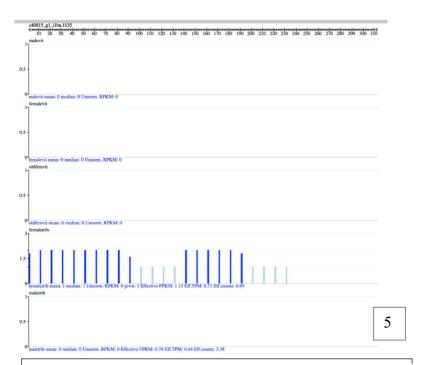


Fig. 5. The expression profile of a mouse aldolase gene. The cDNA was present only in female larvae, and absent from all other stages.

Discussion:

The presence of mouse RNA molecules in the Omaha larval, but not the adult colony, samples strongly suggests that the *tribulatum* larvae ingested mouse tissue. The sequences were not simply conserved genes, as they exactly matched the mouse but had multiple mismatches to human, and no hits to dipterans under the same search conditions.

Mouse sequences were present in both sexes, demonstrating that multiple individuals "ate" mouse. The simplest explanation is the presence of a dead mouse at or upstream from the collecting site. We cannot determine if the larvae accidently collected mouse tissue fragments while filter feeding, or if some larvae deliberately grazed from a mouse body.

Methods and References:

Altschul, S.F., Gish, W., Miller, W., Myers, E.W. & Lipman, D.J. (1990) "Basic local alignment search tool." J. Mol. Biol. 215:403-410.

BLAST searches were performed using NCBI web interface.

Notes for Contributors

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The Simuliid Bulletin / published twice a year / Publisher: Simuliid Research Group, Bradáčova 2, SK-85102 Bratislava, Slovakia, IČO 52544567 / Date of issue: 31.1.2022 (Online) 1.2.2022 (Print) / 1/2022/ Annual volume 29 / Price: free / ISSN 2397-5075 (Online) ISSN: 2397-5067 (Print)/ EV 5887/20.

The Simuliid Bulletin / vychádza dvakrát ročne / Vydavateľ: Simuliid Research Group, Bradáčova 2, SK-85102 Bratislava, Slovakia, IČO: 52544567 / Dátum vydania: 31.1.2022 (online verzia), 1.2.2022 (tlačená verzia) / 1/2022 / 29. ročník / cena: bezplatne / ISSN 2397-5075 (online verzia), ISSN: 2397-5067 (tlačená verzia)/ EV 5887/20.

The Simuliid Bulletin

ISSN 2397-5075 (Online)

Publisher: Simuliid research group

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The Simuliida Bulletin is an informal publication intended to disseminate information about the Simuliidae (Diptera). It is published twice each year on the internet as a downloadable file at URL http://www.simuliid-bulletin.blogspot.com (note the hyphen) or http://www.blackfly.org.uk.

Content covers scientific papers, short research notes, notices and accounts of meetings, and articles of anecdotal or general interest that would not normally be found in international journals. Geographical cover is world-wide. Reports of research carried out by graduates, young scientists and newcomers to the subject are particularly encouraged. It is an ideal medium for offering new ideas and stimulating discussion because of the very short interval between acceptance and publication. Contributions may be accepted up to two weeks before the publication dates at the end of January or July.

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